

Linked Scan Measurements for the Localization of Side-chain Double Bonds in 9,19-Cyclopropano Steroids

I. Kostova,¹ A. Ivanova¹ and H. Budzikiewicz^{2*}

¹ Institute of Organic Chemistry with Centre of Phytochemistry, Bulgarian Academy of Sciences, Sofia 1113, Bulgaria

² Institute of Organic Chemistry, University of Köln, Greinstrasse 4, D-50939 Köln, Germany

9,19-Cyclopropano steroids do not show the typical fragmentation reactions of unsaturated C-17 side-chains in their EI spectra. They can, however, be observed in linked scan measurements of the main fragment arising from ring B cleavage. © 1997 John Wiley & Sons, Ltd.

J. Mass Spectrom. 32, 1317–1319 (1997)

No. of Figures: 2 No. of Tables: 1 No. of Refs: 15

KEYWORDS: 9,19-cyclopropano steroids; side-chain fragmentation; linked scan measurements

INTRODUCTION

Sterols and tetracyclic triterpenoids with a 9,19-cyclopropane ring can be recognized readily from their typical fragmentation behavior.^{1,2} Side-chain unsaturation, which for many steroid results in highly characteristic and pronounced cleavage reactions,³ seems to have only a minor influence or no influence at all on the breakdown pattern. In the course of the structure elucidation⁴ of three terpenoids from *Skimmia wallichii* 1–3 we could show that this problem can be circumvented by linked scan measurements.

EXPERIMENTAL

70 eV mass spectra of 1–3 obtained with an HSQ-30 instrument (Finnigan-MAT, Bremen), *m/z* (%) (relative intensity):

Skimmiwallinin (1): 482 (M^+ , 25), 467 (a, 16), 450 (b, 100), 435 (c, 47), 407 (d, 42), 381 (e, 17), 329 (g, 10), 328 (h, 25), 297 (i, 13), 230 (p, 13), 203 (k, 33), 201 (l, 21), 175 (m, 46), 173 (n, 25).

Skimmiwallichin (2): 496 (M^+ , 21), 481 (a, 13), 464 (b, 100), 449 (c, 33), 421 (d, 29), 395 (e, 17), 342 (h, 21), 297 (i, 10), 230 (p, 17), 203 (k, 23), 201 (l, 13), 175 (m, 33), 173 (n, 15).

* Correspondence to: H. Budzikiewicz, Institut für Organische Chemie, Greinstrasse 4, D-50939 Köln, Germany.

E-mail: h.budzikiewicz@uni-koeln.de.

Contract grant sponsor: Commission of the European Community.

Skimmiwallin (3): 482 (M^+ , 17), 467 (a, 22), 450 (b, 100), 435 (c, 37), 407 (d, 33), 381 (e, 22), 328 (h, 17), 297 (i, 15), 216 (o, 39), 203 (k, 48), 201 (l, 30), 175 (m, 48), 173 (n, 30).

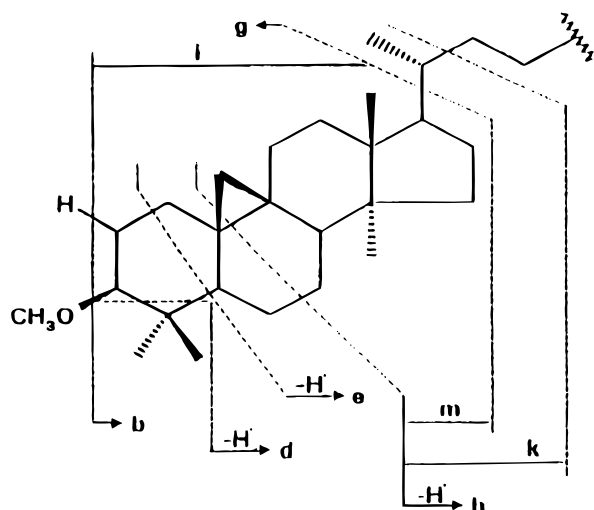
For linked scan ($B/E = \text{const.}$) measurements the gas pressure in the collision cell was adjusted to reduce the intensity of the parent ion to 50% of its original value.

RESULTS AND DISCUSSION

According to Audier *et al.*¹ and Aplin and Hornby,² tetracyclic triterpenoids and steroids with a 9,19-cyclopropane ring undergo diagnostically significant cleavage reactions (for further examples see, Ref. 5). The most important one involves fission of the 9,10-, 9,19- and 5,6-bonds in M^+ with rearrangement of one H, resulting in ion h (Scheme 1), which may subsequently lose the entire side-chain (m; incidentally, parent ion scans of m indicate that this ion can be formed from almost any precursor ion and not—as had been assumed²—only from h). The mass of the ion h is shifted in accordance with the substituents of the side-chain. As parent ion scans show, it originates not only from M^+ but also from $[M - \text{CH}_3]^+$.

Two more fragments seem to be characteristic, namely (in the case of a 3- CH_3O group) $[M - \text{CH}_3\text{OH} - \text{C}_3\text{H}_7]^+$ —possibly d as depicted in Scheme 1—and¹ $[M - \text{CH}_3\text{OH} - \text{C}_5\text{H}_9]^+$ (e in Scheme 1). For other ions—mainly combination losses—see Scheme 1.

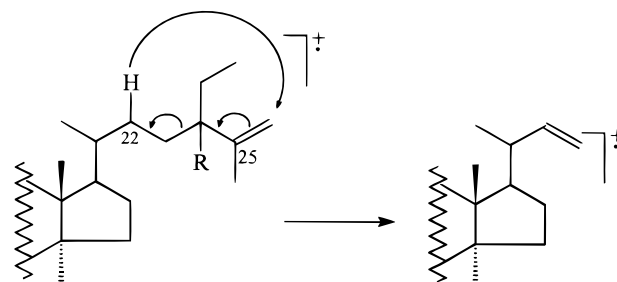
9,19-Cycloterpenoids frequently have an unsaturated side-chain. For compounds without this cyclopropane ring, side-chain double bonds usually give rise to characteristic and abundant fragments.³ In particular, those with a 24-methylene (as in 3) or 24-ethylidene group (24,



Scheme 1. Common fragments of 1–3 (for side-chains see Scheme 2): a $[M - CH_3]^+$, b $[M - CH_3OH]^+$, c $[M - CH_3OH - CH_3]^+$, d $[M - CH_3OH - C_3H_7]^+$, e $[M - CH_3OH - C_5H_9]^+$, g $[M - \text{side-chain}]^+$, h $[M - 154]^+$, i $[b - \text{side-chain}]^+$, k $[M - C-22 \text{ to } C-27]^+$, l $[k - 2H]^+$, m $[h - \text{side-chain}]^+$, n $[m - 2H]^+$.

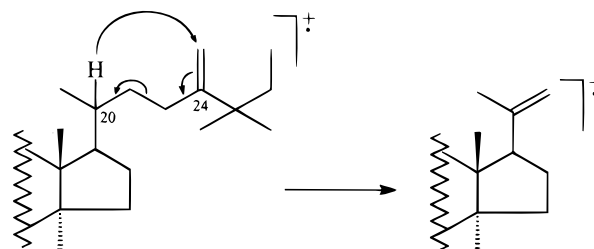
24¹-unsaturation) undergo McLafferty rearrangement with transfer of the C-20 H (see Scheme 2, compound 3). In the case of 3 this process would result in the loss of C₈H₁₆ ($M - 112$; f, m/z 370), absent in the EI spectrum. Even a daughter ion ($B/E = \text{const.}$) scan for $M^{+\cdot}$, which should enhance the relative abundance of McLafferty rearrangement ions, yields m/z 370 only with a relative intensity of 1%.

The side-chain fragmentation induced by a 25,26-double bond is somewhat problematic as far as the literature is concerned. Wyllie and Djerassi³ (and based on their work a review⁶) state that Δ^{25} -compounds also undergo McLafferty rearrangement (see Scheme 2, compounds 1 and 2), quoting a paper by Bergmann *et al.*,⁷ which, however, deals only with 24,24¹-unsaturation (see above). Vulfson and Zaikin⁸ in their review article mention a 24-ethyl- Δ^{25} -steroid which loses not—as expected—C₆H₁₂, but rather C₇H₁₄, and they explain this by the consecutive loss of C₆H₁₁ (allylic cleavage) and CH₃. Further examples of the loss of C_{n+1}H_{2n+2} instead of C_nH_{2n} from Δ^{25} -steroids can be found in the literature (see e.g. Refs 9–11). A more likely explanation would be a shift of the double bond into the tetrasubstituted 24,25-position with subsequent McLafferty rearrangement. 24-Methyl- Δ^{24} -steroids show this rearrangement ion with high intensity,^{7,12} while in the spectra of Δ^{24} -steroids lacking a 24-substituent, where the double bond is not tetrasubstituted, it is missing⁷ (see also Refs 13 and 14). An indication of this double-



- 1, R = CH₃, ($M^{+\cdot}$ m/z 482)
 2, R = C₂H₅, ($M^{+\cdot}$ m/z 496)
 h, R = CH₃ (m/z 328)
 h, R = C₂H₅ (m/z 342)

- j, m/z 384 (1)
 j, m/z 384 (8)
 p, m/z 230 (85)
 p, m/z 230 (100)

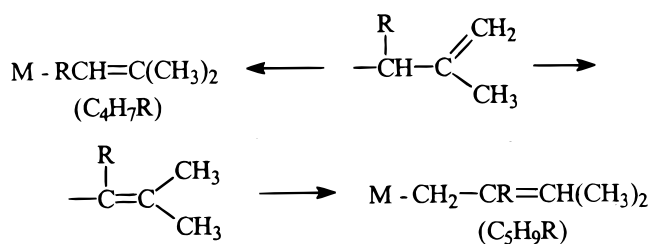


- 3, ($M^{+\cdot}$ m/z 482)
 h, (m/z 328)

- f, m/z 370 (1)
 o, m/z 216 (100)

Scheme 2. Side-chain fragmentation by McLafferty rearrangement (relative intensities in parentheses).

bond migration is the observation that for the sterically hindered 24-isopropyl- Δ^{25} -cholesterol both $[M - C_7H_{14}]^{+\cdot}$ (m/z 328, 17%) and $[M - C_8H_{16}]^{+\cdot}$ (m/z 314, 28%) are present.¹⁵



This double-bond migration should not be possible for 24,24-disubstituted Δ^{25} -steroids, but for the only

Table 1. Daughter ion ($B/E = \text{const.}$) linked scan spectra for h [masses and (in parentheses) relative abundances]

Compound	Parent ion m/z	h - 15	o	p	l	m	n	Other ions
1	328	313 (32)	—	230 (85)	201 (63)	175 (85)	173 (100)	285, 257 (18), (2)
2	342	327 (20)	—	230 (100)	201 (13)	175 (11)	173 (13)	285, 257 (3), (4)
3	328	313 (10)	216 (100)	—	201 (15)	175 (15)	173 (26)	285, 257 (1), (1)

examples without a 9,19-ring known to us where partial mass spectra are reported,⁵ side-chain fragments are not mentioned. In any case, for **1** and **2** in the EI spectra, ions **j** (or their lower homologs) are not observed. In the daughter ion ($B/E = \text{const.}$) linked scans of M^+ they are of low abundance (1% and 8% respectively).

It is therefore important to note that the daughter ion spectra (linked scan $B/E = \text{const.}$) of **h** (Table 1) showed the expected McLafferty fragments (Scheme 2) in high abundance (**1**, **2**: **p**, m/z 230; **3**: **o**, m/z 216). In addition, **m** (m/z 175, loss of the entire side-chain), **n** (m/z 173,

m - 2H) and **l** (loss of C-22 to C-27 + 2H) originating from **h** can be seen. These results show that the B/E linked scan spectra can readily be used for locating sites of unsaturation in the side-chain where the EI spectra do not give the expected results.

Acknowledgement

A fellowship (to I.K.) from the Commission of the European Community is gratefully acknowledged.

REFERENCES

1. H. E. Audier, R. Bengelmans and B. C. Das, *Tetrahedron Lett.* 4341–4347 (1966).
2. R. T. Alpin and G. M. Hornby, *J. Chem. Soc. B* 1078–1079 (1966).
3. S. G. Wyllie and C. Djerassi, *Org. Chem.* **33**, 305 (1968).
4. I. Kostova, M. Simeonov, T. Iossifova, R. Tappe, N. Pardeshi and H. Budzikiewicz, *Phytochemistry* **43**, 643–648 (1996).
5. E. Ritchie, R. G. Senior and W. C. Taylor, *Aust. J. Chem.* **22**, 2371–2387 (1969).
6. H. Budzikiewicz, in *Biochemical Applications of Mass Spectrometry*, edited by G. R. Waller, p. 259. Wiley, New York (1972).
7. J. Bergman, B. O. Lindgren and C. M. Svahn, *Acta Chem. Scand.* **19**, 1661–1666 (1965).
8. N. S. Vulfson and V. G. Zaikin, *Usp. Khim.* **42**, 1379–1414 (1973).
9. W. R. Nes, K. Krevitz, J. Joseph, W. D. Nes, B. Harris, G. F. Gibbons and G. W. Patterson, *Lipids* **12**, 511–527 (1979).
10. V. P. Garg and W. R. Nes, *Phytochemistry* **23**, 2925–2929 (1984).
11. M. D. Greca, P. Monaco and L. Previtera, *Phytochemistry* **28**, 629–631 (1989).
12. J. P. Engelbrecht, B. Tursch and C. Djerassi, *Steroids* 121–126 (1972).
13. F. Borchers, K. Levsen, H. Schwarz, C. Wesdemiotis and R. Wolfschütz, *J. Am. Chem. Soc.* **99**, 1716–1721 (1977).
14. M. Kraft and G. Spittler, *Org. Mass Spectrom.* **2**, 865–876 (1969).
15. W. C. M. Kokke, C. S. Pak, W. Fenical and C. Djerassi, *Helv. Chim. Acta* **62**, 1310–1318 (1979).